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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

	Application No.	Applicant(s)	··
Office Astics Com	10/822,561	DE LA TORRE-BUENO, JOSE	
Office Action Summary	Examiner	Art Unit	
	Colin M. LaRose	2624	
The MAILING DATE of this communication app Period for Reply	ears on the cover sheet with the c	orrespondence addre	PSS
A SHORTENED STATUTORY PERIOD FOR REPLY WHICHEVER IS LONGER, FROM THE MAILING DA - Extensions of time may be available under the provisions of 37 CFR 1.13 after SIX (6) MONTHS from the mailing date of this communication. If NO period for reply is specified above, the maximum statutory period w - Failure to reply within the set or extended period for reply will, by statute, Any reply received by the Office later than three months after the mailing earned patent term adjustment. See 37 CFR 1.704(b).	ATE OF THIS COMMUNICATION 36(a). In no event, however, may a reply be tin will apply and will expire SIX (6) MONTHS from a cause the application to become ABANDONE	N. nely filed the mailing date of this comm D (35 U.S.C. § 133).	
Status			
1) ■ Responsive to communication(s) filed on 24 Ju 2a) ■ This action is FINAL. 2b) ■ This 3) ■ Since this application is in condition for allowar closed in accordance with the practice under E	action is non-final. nce except for formal matters, pro		erits is
Disposition of Claims			
4) Claim(s) 1-18 and 22 is/are pending in the app 4a) Of the above claim(s) is/are withdray 5) Claim(s) is/are allowed. 6) Claim(s) 1-18 and 22 is/are rejected. 7) Claim(s) is/are objected to. 8) Claim(s) are subject to restriction and/or	vn from consideration.		
Application Papers			
9) The specification is objected to by the Examine 10) The drawing(s) filed on is/are: a) access applicant may not request that any objection to the Replacement drawing sheet(s) including the correct 11) The oath or declaration is objected to by the Examine	epted or b) objected to by the drawing(s) be held in abeyance. Se ion is required if the drawing(s) is ob	e 37 CFR 1.85(a). jected to. See 37 CFR	
Priority under 35 U.S.C. § 119			
12) Acknowledgment is made of a claim for foreign a) All b) Some * c) None of: 1. Certified copies of the priority documents 2. Certified copies of the priority documents 3. Copies of the certified copies of the prior application from the International Bureau * See the attached detailed Office action for a list	s have been received. s have been received in Applicat rity documents have been receive u (PCT Rule 17.2(a)).	on No ed in this National Sta	age
Attachment(s) 1) Notice of References Cited (PTO-892) 2) Notice of Draftsperson's Patent Drawing Review (PTO-948) 3) Information Disclosure Statement(s) (PTO/SB/08)	4) Interview Summary Paper No(s)/Mail D 5) Notice of Informal F	ate	
Paper No(s)/Mail Date	6) Other:		

DETAILED ACTION

Amendments and Remarks

1. Applicant's amendments and remarks dated 24 July 2007, have been entered and made of record.

Response to Amendments and Remarks

2. Applicant's arguments have been fully considered but they are not persuasive. In response to applicant's argument that Marcelpoil (7,056,236) and Kotera (4,090,243) are nonanalogous art, it has been held that a prior art reference must either be in the field of applicant's endeavor or, if not, then be reasonably pertinent to the particular problem with which the applicant was concerned, in order to be relied upon as a basis for rejection of the claimed invention. See *In re Oetiker*, 977 F.2d 1443, 24 USPQ2d 1443 (Fed. Cir. 1992).

In this case, Applicant argues not that Marcelpoil and Kotera are not analogous to the claimed invention, but rather that Marcelpoil and Kotera cannot be combined because the field of endeavor of Kotera is not analogous to that of Marcelpoil. Applicant points out various differences between the disclosures of Marcelpoil and Kotera (see Remarks, p. 8).

Notwithstanding these differences, however, Examiner considers the references to be sufficiently closely related for the purposes of § 103 analysis.

Fundamentally, Marcelpoil and Kotera are related to the broad field of image analysis, and more particularly, to the narrower problem of determining the constituent color(s) contained in a sample under examination, in accordance with the claimed invention of "quantifying a color in a sample." In Marcelpoil, the samples under examination are stained or dyed biological cells,

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whereas in Kotera, the samples are painted color prints prepared by artisans. Although the examination of stained cells and the examination of painted prints *per se* cannot be considered related endeavors, both Marcelpoil and Kotera are directed to determining colors present in those samples, and the methods of analysis that each employ—such as defining a matrix based on color values and utilizing an inverse of that matrix based on control measurements as a conversion matrix to calculate the amount of color in the sample (e.g., Kotera, column 4/32 through column 5/66)—are very similar. Accordingly, the disclosures of Marcelpoil and Kotera are considered related at least in the sense that both are directed to analogous problems in the field of image analysis, and both seek solutions to those problems in similar manners.

Claim Objections

3. In view of Applicant's amendments to claims 7 and 16, the previous objections thereto have been withdrawn.

Claim Rejections - 35 USC § 103

- 4. The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:
 - (a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.
- 5. Claims 1-17 and 22 are rejected under 35 U.S.C. 103(a) as being unpatentable over U.S. Patent 7,065,236 by Marcelpoil et al. ("Marcelpoil") in view of U.S. Patent 4,090,243 by Kotera et al. ("Kotera").

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Regarding claims 1 and 10, Marcelpoil discloses a method/program for quantifying color in a sample comprising multiple colors, the method comprising:

measuring a color channel value in a plurality of pixels from a control sample comprising a single color of interest (column 8/14-23: camera 300 captures a color image of a sample 500 – the image having red, green, and blue color channel values);

defining a vector for the control sample, wherein the vector comprises a color channel value present in the control (e.g. the optical density vector OD, given by equations 3-5 or 6-8 in column 11, defines a vector comprising the measured optical densities for the red, green, and blue color channels);

defining a matrix comprising each of the values of each of the color channels (i.e. the matrix formed by the equations associated with the OD vector is defined by equations 21 and 22 in column 14, in order to determine the dye concentrations C based on the known optical densities OD and absorption coefficients ε – see column 14/1-6);

defining a conversion matrix comprising the inverse of the matrix based upon the control measurements (i.e. the conversion matrix denoted by equation 23 in column 14 is defined based upon the measured control optical densities);

measuring color channel values in an image of an experimental sample comprising a plurality of colors of interest, each of the pixels comprising a plurality of color channels (column 16/9-14: an experimental sample having the same dyes uses in the calibration process for determining the conversion matrix is imaged in the same manner as the control sample); and calculating the amount of a color in the experimental sample by converting the channel

values in the experimental sample using the conversion matrix (column 16/14-31: the amount of

color, or concentrations of the dyes, in the experimental sample is determined using the conversion matrix).

However, Marcelpoil seems to only utilize a single control sample and does not appear to disclose or suggest using a "plurality of control samples," as claimed. Accordingly, Marcelpoil does not disclose defining the vector or the matrix on the basis of an "average" of color channel values for a "plurality of control samples."

Kotera discloses a system (figures 1A and 1B) for characterizing the colors of a color sample that is very similar to that of Marcelpoil and involves the same concepts of deriving an inverse matrix of mean color intensity values (column 5/1-35) and using the inverse matrix to ascertain the colors of an experimental sample (column 5/58-66). In particular, Kotera teaches that control color prints C1 ... Cn, as shown in figure 2, are imaged, and the measured colors and used to determine the conversion matrix. Kotera teaches that each of these control color prints contain a plurality of "elemental areas," which essentially correspond to sub-areas within the larger print area, and the elemental areas are each "microscopically" imaged to generate representative intensity signals thereof (column 1/59 through column 2/3). The mean intensity value μ of each spectral (i.e. color) component for each control color print corresponds to the representative intensity signal of a per-unit area (column 2/18-27). The set of mean values μ , in conjunction with the set of representative intensity signals, is utilized to obtain the conversion matrix that is used to ascertain the color C_i of a given experimental sample, expressed as the probability P(C_i) that the ascertained color corresponds to the i-th control print (see column 2/22-39 and column 3/57 et seq.).

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It would have been obvious to one of ordinary skill in the art at the time of the invention to modify Marcelpoil by Kotera to achieve the claimed invention by measuring a plurality of control samples and defining the vector and the matrix on the basis of an average of each color channel present in the control samples, since Kotera shows that it was conventional to microscopically scan a control color print on the basis of a plurality of "elemental" sample areas and utilize the average of those sample areas as a basis for deriving a conversion matrix, as explained above. Such a modification would achieve substantially the same results as achieved by Marcelpoil since Marcelpoil's control samples are analyzed in a region where two or three dyes are mixed to produce an area of uniform color.

Regarding claims 2 and 11, Marcelpoil discloses the color channels comprise red, green, and blue (see figure 1).

Regarding claims 3 and 12, the combination of Marcelpoil and Kotera discloses each control is stained with a single staining reagent to generate a color of interest (column 9/13-17 and column 10/63 et seq. of Marcelpoil: the control is stained with a plurality of staining reagents, including a single marker dye that is used to generate a color of interest).

Regarding claims 4 and 13, Marcelpoil discloses that the experimental sample is stained with a plurality of stains to generate a plurality of colors of interest (column 10/63 et seq.: the sample is stained with e.g. a marker dye and a counterstain).

Regarding claims 5 and 14, Marcelpoil discloses that the number of stains in a experimental sample are less than or equal to the number of color channels (column 16/14-31: concentrations of 3 dyes are determined – with there being 3 color channels).

Regarding claims 6 and 15, Marcelpoil suggests that an image of the experimental sample can be displayed as a monochrome image (see e.g. equation 24, column 16, which quantifies the black and white pixel intensities for the experimental sample image).

Regarding claims 7 and 16, Marcelpoil does not expressly disclose setting all but one of the color channel values to zero in order to determine the amount of a single color in the experimental sample, as claimed, however, such a limitation would have been exceedingly well-known and obvious to those skilled in the art in view of the fact that each of the color channels for an RGB image independently quantify the amount of a single color – red, green, or blue – present in an image, and e.g. the values of red and green channels have no bearing on how much "blue" is exhibited by an image of the sample.

Regarding claims 8 and 17, Marcelpoil discloses rendering a digital display of the experimental sample (i.e. displayed on the computer screen, as shown in figures 1 and 2).

Regarding claim 9, the combination of Marcelpoil and Kotera teaches the computer implemented method of claim 1 (i.e. both Marcelpoil's and Kotera's methods are implemented via a computer).

Regarding claim 22, Marcelpoil discloses that the computer-implemented method can be automated (see column 20/2-14).

6. Claim 18 is rejected under 35 U.S.C. 103(a) as being unpatentable over U.S. Patent 7,065,236 by Marcelpoil et al. ("Marcelpoil") in view of U.S. Patent 4,090,243 by Kotera et al. ("Kotera") and U.S. Patent Application Publication 2004/0114227 by Henderson et al. ("Henderson").

Regarding claim 18, Marcelpoil discloses a machine vision system (figures 1 and 2) for automated analysis of a biological sample on a slide comprising:

a system processor (i.e. computer 350 includes a processor);

a computer program on computer readable medium (column 20/2-14), the computer program comprising an image algorithm comprising instructions to cause the computer to:

measure a color channel value in a plurality of pixels from a control sample comprising a single color of interest (column 8/14-23: camera 300 captures a color image of a sample 500 – the image having red, green, and blue color channel values);

define a vector for the control sample, wherein the vector comprises a color channel value present in the control (e.g. the optical density vector OD, given by equations 3-5 or 6-8 in column 11, defines a vector comprising the measured optical densities for the red, green, and blue color channels);

define a matrix comprising each of the values of each of the color channels (i.e. the matrix formed by the equations associated with the OD vector is defined by equations 21 and 22 in column 14, in order to determine the dye concentrations C based on the known optical densities OD and absorption coefficients ε – see column 14/1-6);

define a conversion matrix comprising the inverse of the matrix based upon the control measurements (i.e. the conversion matrix denoted by equation 23 in column 14 is defined based upon the measured control optical densities);

measure color channel values in an image of an experimental sample comprising a plurality of colors of interest, each of the pixels comprising a plurality of color channels (column

the conversion matrix); and

2);

16/9-14: an experimental sample having the same dyes uses in the calibration process for determining the conversion matrix is imaged in the same manner as the control sample); and calculate the amount of a color in the experimental sample by converting the channel values in the experimental sample using the conversion matrix (column 16/14-31: the amount of color, or concentrations of the dyes, in the experimental sample is determined using

output the amount of color in the experimental sample (column 17/1-19); a monitor in operable communication with the computer (as shown in figure 1); an input device in connection with the computer (e.g. keyboard or mouse shown in figure

an optical imaging system (video microscopy system 100) in operable communication with the computer, comprising:

a movable stage (column 18/59-63);

an identification member (column 17/28-45: identification marks produced by an operator);

an optical sensing member (camera 300) in optical communication with the stage configured to acquire an image at a location on a slide and in electrical communication with the processor;

a storage member for storing the location of a candidate object or area of interest (column 17/20-64 and column 19/28-46: the memory of the computer 350 is used to store images containing markings that indicate the locations of areas of interest); and

a storage device for storing each image (column 19/22-32).

However, Marcelpoil seems to only utilize a single control sample and does not appear to disclose or suggest using a "plurality of control samples," as claimed. Accordingly, Marcelpoil does not disclose defining the vector or the matrix on the basis of an "average" of color channel values for a "plurality of control samples."

Kotera discloses a system (figures 1A and 1B) for characterizing the colors of a color sample that is very similar to that of Marcelpoil and involves the same concepts of deriving an inverse matrix of mean color intensity values (column 5/1-35) and using the inverse matrix to ascertain the colors of an experimental sample (column 5/58-66). In particular, Kotera teaches that control color prints C1 ... Cn, as shown in figure 2, are imaged, and the measured colors and used to determine the conversion matrix. Kotera teaches that each of these control color prints contain a plurality of "elemental areas," which essentially correspond to sub-areas within the larger print area, and the elemental areas are each "microscopically" imaged to generate representative intensity signals thereof (column 1/59 through column 2/3). The mean intensity value μ of each spectral (i.e. color) component for each control color print corresponds to the representative intensity signal of a per-unit area (column 2/18-27). The set of mean values μ , in conjunction with the set of representative intensity signals, is utilized to obtain the conversion matrix that is used to ascertain the color C_i of a given experimental sample, expressed as the probability P(C_i) that the ascertained color corresponds to the i-th control print (see column 2/22-39 and column 3/57 et seq.).

It would have been obvious to one of ordinary skill in the art at the time of the invention to modify Marcelpoil by Kotera to measure a plurality of control samples and define the vector

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and the matrix on the basis of an average of each color channel present in the control samples, as claimed, since Kotera shows that it was conventional to microscopically scan a control color print on the basis of a plurality of "elemental" sample areas and utilize the average of those sample areas as a basis for deriving a conversion matrix, as explained above. Such a modification would achieve substantially the same results as achieved by Marcelpoil since Marcelpoil's control samples are analyzed in a region where two or three dyes are mixed to produce an area of uniform color.

In addition, Marcelpoil discloses that the microscope may include one or more robotic components (column 18/59-63) but does not appear to disclose an automated loading and unloading member for loading and unloading of a slide, as claimed.

Henderson discloses an automated slide loader for use with a microscope. In particular, Henderson teaches that it is advantageous to provide an apparatus that automatically loads and unloads slides to and from a microscope. It would have been obvious to one of ordinary skill in the art at the time of the invention to modify Marcelpoil and Kotera to achieve the claimed invention by including an automatic slide loader/unloader, as claimed, since automating a manual procedure has been judicially recognized as per se obvious. See *In re Venner*, 262 F.2d 91, 95, 120 USPQ 193, 194 (CCPA 1958).

Conclusion

7. THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the mailing date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Colin M. LaRose whose telephone number is (571) 272-7423. If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Brian Werner, can be reached on (571) 272-7401. The fax phone number for the organization where this application or proceeding is assigned is (571) 273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000. Any inquiry

of a general nature or relating to the status of this application or proceeding can also be directed to the TC 2600 Customer Service Office whose telephone number is (571) 272-2600.

Colin M. LaRose Group Art Unit 2624

9 October 2007